

Comparative Evaluation of Antimicrobial Efficacy of Phytomedicinal Extracts and Chemical Irrigant against *Enterococcus faecalis* and *Candida albicans*: In vitro study

Kapil D. Wahane^{1*} Sweety Kumari² Sadashiv G. Daokar³ Kalpana S. Pawar⁴ Jyoti Magare⁵ Kalyani S. Kagne⁶

¹Reader & PG Guide, Department of Conservative Dentistry and Endodontics, C.S.M.S.S Dental College and Hospital, Aurangabad, Maharashtra, India.

²PG Student, Department of Conservative Dentistry and Endodontics, C.S.M.S.S Dental College and Hospital, Aurangabad, Maharashtra, India.

³Professor & Head, Department of Conservative Dentistry and Endodontics, C.S.M.S.S Dental College and Hospital, Aurangabad, Maharashtra, India.

⁴Professor & PG Guide, Department of Conservative Dentistry and Endodontics, C.S.M.S.S Dental College and Hospital, Aurangabad, Maharashtra, India.

⁵Reader, Department of Pathology and Microbiology, C.S.M.S.S Dental College and Hospital, Aurangabad, Maharashtra, India.

⁶PG Student, Department of Conservative Dentistry and Endodontics, C.S.M.S.S Dental College and Hospital, Aurangabad, Maharashtra, India.

ABSTRACT

Background: To compare and evaluate the antimicrobial activity of phytomedicinal extracts and chemical irrigant against *E. faecalis* and *C. albicans* using disc diffusion method.

Materials and Methods: For the study, microbial strain of *Enterococcus faecalis* and *Candida albicans* were used. Four experimental groups were divided. These were as follow- Group I- Garlic Extracts, Group II- Ginger extracts, Group III- 5.25% Sodium Hypochlorite and Group IV- 2 % Chlorohexidine Gluconate. For the agar disc diffusion test, Petri plates of Muller Hinton agar were inoculated with 0.1 ml of the microbial suspensions. Subsequently, the paper discs of four groups were placed on the Muller Hinton agar plate. After 24 hours, the diameter of inhibition zone was measured around the papers discs containing the substances.

Results: Statistically significant difference ($p < 0.05$) was found between the groups against *E. faecalis* and *C. albicans*. Order of antimicrobial activity is Garlic > 2 % Chlorohexidine gluconate > 5.25% Sodium Hypochlorite > Ginger.

Conclusions: Garlic extract is advantageous in cases of secondary endodontic infections which are dominated by organisms like *E. faecalis* and *C. albicans*.

Keywords: *Enterococcus faecalis*, *Candida albicans*, disc diffusion, ginger extract, garlic extract.

INTRODUCTION

One of the key reasons behind the success of root canal treatment is the chemo-mechanical preparation which aids in removal of remaining tissue, various microbes and debris from the complex root canal areas and allow the canals to be obturated to achieve a fluid tight seal which will

prevent future ingress of microbes into the canals⁽¹⁾. Although a plethora of microbes have been found to reside in oral cavity, the lower oxygen potential inside root canals and nutrient availability leads to colony formation of facultative anaerobes predominantly.

Received: Nov. 15, 2019; Accepted: Dec. 17, 2019

*Correspondence Dr. Kapil D. Wahane.

Department of Conservative Dentistry and Endodontics, C.S.M.S.S Dental College and Hospital, Aurangabad, Maharashtra, India.

Email: drsweetylohani@gmail.com

Enterococcus faecalis, a facultative anaerobe found in 4 to 40% of primary endodontic infections, survives unaided inside canals and periapical area and multiply by genetic polymorphism, causes triggering infection and local bone resorption⁽²⁾. Because of its virulence factors like aggregation substance, cytolysin and lytic enzymes can alter host responses and hence it is able to thrive better than other microbes and is the strain isolated after unsuccessful endodontic treatment⁽³⁾. Moreover, they invade dentinal tubules and adhere to collagen due to their ability to form biofilm and therefore it may act as a nidus for recurrence of infection in failed root canal treated teeth⁽⁴⁾.

Recent studies have shown that fungi also cause root canal infections. Fungi have been detected in root canals with primary and secondary endodontic infections⁽⁴⁾ abscesses or cellulitis of endodontic origin, and Periapical lesions in asymptomatic teeth⁽⁵⁾.

Candida albicans is the most commonly isolated, potentially pathogenic yeast species from infected root canals⁽⁵⁾. *Candida* is a versatile micro-organism, each form of growth depends on environmental conditions such as the pH level, temperature, and nutritional source⁽⁶⁾. Furthermore, it also exhibits a variety of virulence factors such as adherence, thigmotropism and phenotypic switching and secretes a degenerative enzyme "aspartyl protease" that degrades the dentinal collagen⁽⁷⁾.

Because of these unique qualities, able to survive in the harsh ecologic environment of the root canal, *E. faecalis* and *C. albicans* choose as test micro-organism.

Hence, to attain lasting successful root canal treatment, it necessitates a complete disinfection of root canal and limiting the recurrence of infection. Use of irrigants to thoroughly clean root canal systems during instrumentation is central to successful endodontic treatment. Irrigation is complementary to instrumentation and facilitates the removal of pulp tissues and/or microorganisms. In addition to providing mechanical flushing action, an irrigant should exert microbicidal effects without damaging periradicular tissues⁽⁸⁾.

Sodium hypochlorite (NaOCl) is a potent irrigant with a broad-spectrum antimicrobial activity that

causes the breakdown of proteins and amino acids through its "free chlorine release." It has the potential to dissolve the necrotic pulp tissue and organic debris⁽⁹⁾.

Chlorohexidine gluconate (CHX) is also a broad-spectrum antimicrobial agent with a wide range of action. CHX has a unique feature in that dentin medicated with it acquires antimicrobial substantivity. The positively charged ions released by CHX can adsorb into dentin and prevent microbial colonization for some time beyond the time of the medicament⁽¹⁰⁾. Side effects of synthetic drugs have prompted researchers to search for herbal alternatives. Pharmacological studies have acknowledged the value of medicinal plants as potential sources of bioactive compounds⁽¹¹⁾. Some natural plant extracts have antimicrobial and therapeutic properties, suggesting their potential use as endodontic irrigants or intracanal medications⁽¹²⁾. In addition to antimicrobial effects, herbal irrigants are safe and nontoxic to host tissues.

The aim of the present study was to evaluate the antimicrobial efficacy of phytomedicinal extracts of garlic and ginger with chemical irrigant of 5.25% sodium hypochlorite (NaOCl) and 2% chlorhexidine gluconate (CHX) against *E. faecalis* and *C. albicans*.

Aim: To evaluate the antimicrobial efficacy of phytomedicinal extracts of Garlic and Ginger as endodontic irrigants with chemical irrigants of 5.25% NaOCl and 2% Chlorohexidine gluconate against *Enterococcus faecalis* and *Candida albicans*.

MATERIALS AND METHODS

The following materials were used in the study:

Extract of garlic, ginger, 2% Chlorohexidine Gluconate and 5.25% Sodium hypochlorite, *Candida albicans* strain, *E. faecalis* strain, Chromagar, Mitis Salivarius Agar, Mueller Hinton Agar, sterile test-tubes and sterile droppers, Whatman filter paper no.1.

Preparation of extracts

Preparation of garlic and ginger extract

Fresh natural garlic and ginger were purchased from local market. The cloves were separated from



Fig 3: Growth of *C. albicans* on Chromagar.

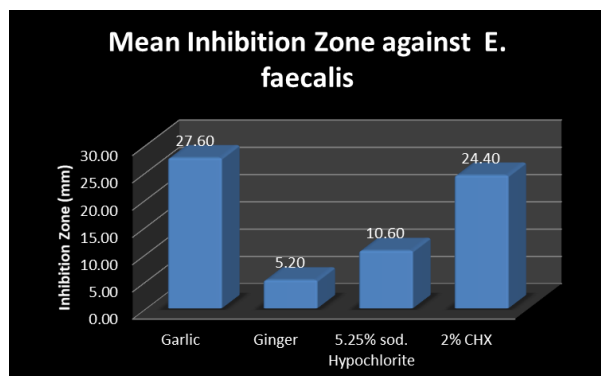


A petri dish containing a bacterial culture on a blue agar surface. The culture shows a large, dark, circular colony with a smaller, more defined colony inside it. Handwritten markings "sb" and "P" are visible on the agar surface.

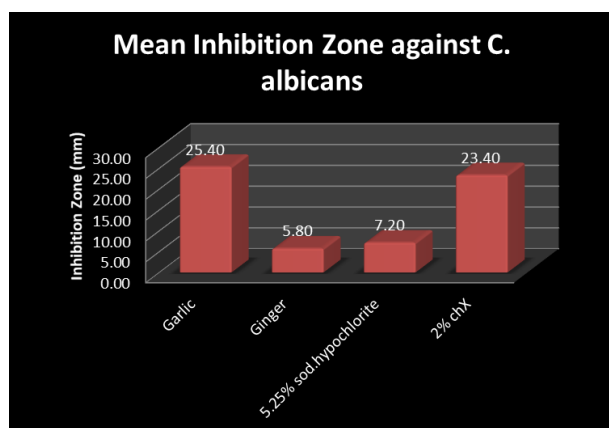
A petri dish containing a bacterial culture on a nutrient agar plate. The plate is divided into four quadrants by a black 'X' mark. The quadrants are labeled: 'Sod. Hypothese' (top-left), 'Glu' (top-right), 'Glu' (bottom-left), and 'Glu' (bottom-right). The growth is visible as white, fuzzy colonies on the surface of the agar.

Fig 5: Inhibition Zone of study group against *C. albicans*.

Graph 1: Mean zone of inhibition of all group against *E. faecalis*.



Graph 2: Mean zone of inhibition of all group against *C. albicans*.



garlic and peeled to obtain the edible portion. Fifty gram of the edible portion was chopped and homogenized in 100 ml of autoclaved water in a blender. The homogenized was then filtered by passing through a 25µm pore size filter to give a crude aqueous extract of 500mg of extracts/ml. This was collected in a sterile screw cork bottle and stored at 4°C until used.

Microbial strains

Two strains including *Candida albicans* and *Enterococci faecalis* were obtained from the Microbial Culture Collection Bank, Department of Pathology and Microbiology. The *Candida* strains were maintained on Sabouraud's dextrose agar slants and *E. faecalis* were maintained on Brain Heart Infusion. The culture was kept under refrigerated conditions and was sub cultured every 15 days. After that, growth of pure colonies of *E. faecalis* on Mitis Salivarius agar with 1% potassium

tellurite and *C. albicans* on Chromagar were maintained.

Preparation of antimicrobial disc

Whatman filter paper no.1 was used for preparation of antimicrobial disc. The paper disc were prepared and kept in hot air oven at 160°C for 1 hr for sterilization. The antimicrobials agents that are to be used are impregnated on sterile filter paper disc. Prepared antimicrobial disc divided into four group.

Group I- Garlic Extracts

Group II- Ginger Extracts

Group III- 5.25% Sodium Hypochlorite

Group IV – 2% Chlorohexidine Gluconate

These disc were kept for drying in oven. Prepared antimicrobial discs were stored in refrigerators.

Preparation of microbial inoculum

The density of selected organisms was adjusted equal to that of the 0.5 McFarland standards by adding them to BHI broth for *E. faecalis* and SDA for *C. albicans*. A 24 h old culture was used for the preparation of bacterial suspension. McFarland standards were used as a reference to adjust the turbidity of microbial suspension so that the number of microorganisms would be within a given range.

Antimicrobial susceptibility test

Disc diffusion method was used to conduct the antimicrobial susceptibility test on Muller Hinton agar by Kirby Bauer Method. The agar media plates were inoculated with the organisms by even streaking of the swab over the entire surface of the Muller Hinton agar plate 3 times, rotating the plate approximately 60° after each application to ensure an even distribution of the inoculums. Prepared antimicrobial bio disc were placed equidistance on agar plate. The plates were maintained for 1 h at room temperature, and then the plates, thus prepared were incubated for 24 h at 37°C. The zone of inhibition was measured and expressed in millimetres. Experiments were performed 5 times and mean of the zone of inhibition was recorded in mm. The mean and standard deviation of the diameter of inhibition zones were calculated.

Table 1: Mean, Standard deviation, Standard error of inhibition zone against *E. faecalis*.

Group	Mean	Std. Deviation	Std. Error	95% Confidence Interval for Mean	
				Lower Bound	Upper Bound
Garlic	27.60	1.673	.748	25.52	29.68
Ginger	5.20	1.483	.663	3.36	7.04
5.25 % Sodium Hypochlorite	10.60	1.342	.600	8.93	12.27
2% CHX	24.40	1.517	.678	22.52	26.28
Total	16.95	9.660	2.160	12.43	21.47

Table 2: Mean, Standard deviation, Standard error of inhibition zone against *C. albicans*.

Group	Mean	Std. Deviation	Std. Error	95% Confidence Interval for Mean	
				Lower Bound	Upper Bound
Garlic	25.40	.894	.400	24.29	26.51
Ginger	5.80	.837	.374	4.76	6.84
5.25% Sodium Hypochlorite	7.20	2.168	.970	4.51	9.89
2% CHX	23.40	.894	.400	22.29	24.51
Total	15.45	9.305	2.081	11.10	19.80

STATISTICAL ANALYSIS

All data were analysed by using IBM SPSS-20 software. The data was interpreted at a confidence interval of 95%. Analysis of variance (ANOVA) was used to compare the antimicrobial activity of herbal extracts with chemical irrigant. Significance level was set at $P \leq 0.05$.

RESULTS

The mean values of inhibition zone were obtained in the groups by zone scale. Mean zone of inhibition were presented in Table 1 and Graph 1 against *E. faecalis* and Table 2 and Graph 2 against *C. albicans*. Order of antimicrobial activity is Garlic > 2 % chlorohexidine gluconate > 5.25 % sodium hypochlorite > ginger against *E. faecalis* and *C. albicans*. The antimicrobial activity of garlic extract is more effective against *E. faecalis* and *C. albicans* than the 2 % chlorohexidine gluconate and the difference being statistically significant. The

antimicrobial activity of 2% Chlorohexidine gluconate is more than sodium hypochlorite but less than garlic extract against *E. faecalis* and *C. albicans*. The antimicrobial property of 5% sodium hypochlorite and ginger were found least in both group against *E. faecalis* and *C. albicans*.

DISCUSSION

Primary root canal infections consist of different types of microbes, mostly consisting of obligate anaerobes⁽¹³⁾. It is believed that both mechanical enlargement and copious irrigation of root canals aids in maximum removal of microorganisms as possible^(14,15). Various studies show that the obligate anaerobes are easier to eliminate through endodontic procedures than facultative anaerobes (isolated in 19% to 22% of cases); the latter prove more resistant to chemo-mechanical procedures. Facultative anaerobic micro-organisms such as *Enterococcus faecalis*, *Staphylococcus aureus*, and *Candida albicans* are considered to have the highest

resistance in the oral cavity, with the potential to cause failure of root canal treatment⁽¹⁶⁾. In particular *Enterococcus faecalis* and *C. albicans* have gained attention in the endodontic literature, as associated with therapy resistant apical periodontitis. Therefore, *E. faecalis* and *C. albicans* choose for study purpose as test organism.

Irrigation is important for root canal treatment as it assists in removing bacteria and debris within the intricacies of the root canal. An irrigant should impart mechanical, microbicidal properties, be biocompatible and aid in dissolving organic tissues without damaging the periradicular tissues if extrusion of irrigant occurs into the periodontium.

Sodium hypochlorite (NaOCl) is the most popular irrigant used during instrumentation. It is a nonspecific proteolytic agent with wide-ranging activities against endodontic microorganisms. Higher the concentration of NaOCl, better are the antibacterial and tissue dissolution properties, but at the same time, more are the chances of tissue reaction. Hence, 5.25% NaOCl is clinically more acceptable and has been used in this study.

Chlorohexidine gluconate (CHX) is widely used for disinfection in dentistry because of its good antimicrobial activity. CHX has been shown to have long-term antimicrobial properties because of its unique ability to bind to hydroxyapatite. It was proved that 2% CHX liquid was more potent than 1% and 0.2% liquid against selected microorganisms⁽¹⁶⁾. Hence, in this study, 2% CHX liquid was used.

A wide variety of synthetic antimicrobial agents have been used over the years as endodontic irrigants. Because of the increased antibiotic resistance to these antimicrobial agents, toxic and harmful side effects of few common antibacterial agents, there is a need for alternative agents which are affordable, nontoxic, and effective. It has been found that natural plant extracts could be used as effective endodontic irrigants⁽¹⁷⁾.

Phytochemicals derived from plant products serve as a prototype to develop less toxic and more effective medicines in controlling the growth of micro-organism^(18,19). These compounds have significant therapeutic application against human pathogens including bacteria, fungi or virus.

Numerous studies have been conducted with the extracts of various plants, screening antimicrobial activity as well as for the discovery of new antimicrobial compounds⁽²⁰⁾.

Garlic has antimicrobial⁽²¹⁾ anticancer⁽²²⁾ and antiplatelet activity⁽²³⁾ **Eswar et al.** investigated the efficacy of Garlic against *E. faecalis*. Garlic showed better antibacterial efficacy due to the presence of allicin, an organosulphur compound in garlic^(24,25).

Ginger-It belongs to *Zingiberaceae* family and it has also been used as a medicine from Vedic days due to its antibacterial and antifungal properties. **Maekawa et al**⁽²⁶⁾ evaluated the antimicrobial efficacy of Glycolic propolis, ginger extract, Ca(OH)₂, CHX and their combinations against *E. faecalis*, *C. albicans*, *Escherichia coli* and endotoxins in root canals. Ginger contains flavonoids, It was observed that this extract was effective in eliminating the microorganisms from the root canals⁽²⁷⁾.

In the present study, garlic extract showed maximum antimicrobial efficacy against *E. faecalis* and *C. albicans*. Garlic contains at least 33 sulphur compounds, several enzymes, amino acids, and minerals such as selenium. It contains a higher concentration of sulphur compounds than any other *Allium* species⁽²⁸⁾. Allicin in garlic extracts have been found to be effective as an antifungal, antibacterial, antiviral and anti-parasitic agent. The present study was in with **Suresh Pandey et al**⁽²⁹⁾ also concluded that Garlic extract is advantageous in cases of secondary endodontic infections which are dominated by organisms like *E. faecalis* and *C. albicans*.

In our study, 2% CHX showed better antibacterial efficacy compared to sodium hypochlorite and ginger. The possible reasons might be due to bactericidal dosage of 2% CHX and increased diffusion of the medicament into the test media. Its efficacy is because of the interaction of the positive charge of the molecule with the negatively charged phosphate groups on microbial cell walls, which alters the cells osmotic equilibrium. This increases the permeability of the cell wall, allowing the CHX molecule to penetrate into the bacteria. Damage to this delicate membrane is followed by leakage of intracellular constituents, particularly phosphate entities such as adenosine triphosphate and nucleic acids. As a consequence, the cytoplasm becomes

congealed, with resultant reduction in leakage; thus, there is a biphasic effect on membrane permeability⁽³⁰⁾. The result of this present study was similar to that of **Krithikadatta et al**⁽³¹⁾ **Gomes et al**⁽³²⁾ and **Bhardwaj et al**⁽³³⁾.

In our study sodium hypochlorite and ginger shows less inhibition zone, might be due to experimental methods, biological indicators and exposure time.

In this study, the antimicrobial activity of herbal extracts of Garlic had more inhibition zone than ginger. However, several disadvantages of herbal irrigants like fresh solutions has to be prepared each time, the unacceptable odour and taste, short shelf life have to be overcome. The smell and taste of the herbal irrigants has to be modified by adding flavouring agents to make it palatable and acceptable by the patient. More research for prolonging the shelf life of

these irrigants has to be done so that these irrigants are more widely accepted.[40]

CONCLUSION

Under the conditions of the present study and within its limitations, it can be concluded that:

1. With ever increasing resistance to synthetic drugs and typical features of resistant of antimicrobial agents of *E.faecalis* and *C. albicans*, herbal extracts can be an alternative option provided all the ideal properties of an irrigant are satisfied. Garlic extracts have antimicrobial properties than ginger.
2. 2% Chlorohexidine solution is a better endodontic irrigant with maximum antimicrobial potency compared to 5.25% sodium hypochlorite and ginger.
3. Agar diffusion test have its own drawbacks and cannot simulate the *in vivo* conditions which might vary the results obtained in the present study. Further research is needed to investigate the availability of aqueous and other properties of garlic that has more adequate properties for irrigant. Although the *in vitro* test methods are helpful in conducting and evaluating the antibacterial efficacy of various irrigants, a detailed and standardised procedure is

required to enhance the reproducibility among different researchers in future.

CONFLICTS OF INTEREST

The authors declare they have no potential conflict of interests regarding this article.

REFERENCES

1. Tronstad L, Sunde PT. The evolving new understanding of endodontic infections. *Endodontic Topics*. 2003 Nov;6(1):57-77.
2. Bazvand L, Aminozarbian MG, Farhad A, Noormohammadi H, Hasheminia SM, Mobasherizadeh S. Antibacterial effect of triantibiotic mixture, chlorhexidine gel, and two natural materials Propolis and Aloe vera against *Enterococcus faecalis*: An ex vivo study. *Dental research journal*. 2014 Jul;11(4):469-474
3. Dumani A, Yoldas O, Yilmaz S, Koksall F, Kayar B, Akcimen B, Seydaoglu G. Polymerase chain reaction of enterococcus faecalis and candida albicans in apical periodontitis from Turkish patients. *Journal of clinical and experimental dentistry*. 2012 Feb;4(1):e34-e39.
4. Eswar K, Venkateshbabu N, Rajeswari K, Kandaswamy D. Dentinal tubule disinfection with 2% chlorhexidine, garlic extract, and calcium hydroxide against *Enterococcus faecalis* by using real-time polymerase chain reaction: In vitro study. *Journal of conservative dentistry: JCD*. 2013 May;16(3):194-198.
5. Baumgartner JC, Watts CM, Xia T. Occurrence of *Candida albicans* in infections of endodontic origin. *Journal of endodontics*. 2000 Dec 1;26(12):695-8.
6. Nair PR, Sjögren U, Krey G, Kahnberg KE, Sundqvist G. Intraradicular bacteria and fungi in root-filled, asymptomatic human teeth with therapy-resistant periapical lesions: a long-term light and electron microscopic follow-up study. *Journal of endodontics*. 1990 Dec 1;16(12):580-588.
7. Şen BH, Safavi KE, Spångberg LS. Growth patterns of *Candida albicans* in relation to radicular dentin. *Oral Surgery, Oral Medicine,*

- Oral Pathology, Oral Radiology, and Endodontology. 1997 Jul 1;84(1):68-73.
8. Becker TD, Woollard GW. Endodontic irrigation. General dentistry. 2001;49(3):272-276.
 9. Mohammadi Z. Sodium hypochlorite in endodontics: an update review. International dental journal. 2008 Dec;58(6):329-341.
 10. Mohammadi Z, Abbott PV. The properties and applications of chlorhexidine in endodontics. International endodontic journal. 2009 Apr;42(4):288-302.
 11. Nabavizadeh M, Abbaszadegan A, Gholami A, Sheikhi R, Shokouhi M, Shams MS, Ghasemi Y. Chemical constituent and antimicrobial effect of essential oil from *Myrtus communis* leaves on microorganisms involved in persistent endodontic infection compared to two common endodontic irrigants: An in vitro study. Journal of conservative dentistry: JCD. 2014 Sep;17(5):449-453.
 12. Vinothkumar TS, Rubin MI, Balaji L, Kandaswamy D. In vitro evaluation of five different herbal extracts as an antimicrobial endodontic irrigant using real time quantitative polymerase chain reaction. Journal of conservative dentistry: JCD. 2013 Mar;16(2):167-170.
 13. Sundqvist G. Taxonomy, ecology, and pathogenicity of the root canal flora. Oral Surgery, Oral Medicine, Oral Pathology. 1994 Oct 1;78(4):522-530.
 14. Lin LM, Skribner JE, Gaengler P. Factors associated with endodontic treatment failures. Journal of endodontics. 1992 Dec 1;18(12):625-627.
 15. Eldeniz AU, Hadimli HH, Ataoglu H, Ørstavik D. Antibacterial effect of selected root-end filling materials. Journal of endodontics. 2006 Apr 1;32(4):345-349.
 16. Vianna ME, Gomes BP, Berber VB, Zaia AA, Ferraz CC, de Souza-Filho FJ. In vitro evaluation of the antimicrobial activity of chlorhexidine and sodium hypochlorite. Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology, and Endodontology. 2004 Jan 1;97(1):79-84.
 17. Kelmanson JE, Jäger AK, van Staden J. Zulu medicinal plants with antibacterial activity. Journal of Ethnopharmacology. 2000 Mar 1;69(3):241-246.
 18. Ahmad I, Beg AZ. Antimicrobial and phytochemical studies on 45 Indian medicinal plants against multi-drug resistant human pathogens. Journal of ethnopharmacology. 2001 Feb 1;74(2):113-123.
 19. Guleria S, Kumar A. Antifungal activity of some Himalayan medicinal plants using direct bioautography. Journal of cell and molecular Biology. 2006 Jun 1;5:95-98.
 20. Zakaria Z, Sreenivasan S, MOHAMAD M. Antimicrobial Activity of Piper ribesoides Root Extract Against *Staphylococcus aureus*. Journal of Applied Biological Sciences. 2007 Sep 1;1(3):87-90
 21. Shetty S, Thomas B, Shetty V, Bhandary R, Shetty RM. An in-vitro evaluation of the efficacy of garlic extract as an antimicrobial agent on periodontal pathogens: A microbiological study. Ayu. 2013 Oct;34(4):445-451
 22. Milner JA. Garlic: its anticarcinogenic and antitumorigenic properties. Nutrition reviews. 1996 Nov;54(11 Pt 2):82-86.
 23. Vilahur G, Badimon L. Antiplatelet properties of natural products. Vascular Pharmacology. 2013 Sep 1;59(3-4):67-75.
 24. Feldberg R.S., Chang S.C., Kotik A.N., Nadler M., Neuwirth Z., Sundstrom D.C., Thompson N.H. *In vitro* mechanism of inhibition of bacterial cell growth by allicin. Antimicrob. Agents Chemother. 1988;32(12):1763-1768
 25. Panchajanya S. An In-vitro Evaluation Of Antibacterial Activity Of Medicinal Plants And Calcium Hydroxide Against *Enterococcus Faecalis* By Modified Direct Contact Test. Indian Journal of Dental Sciences. 2014 Jun 1;6(2).
 26. Maekawa L.E., Valera M.C., Oliveira L.D., Carvalho C.A., Camargo C.H., Jorge A.O. Effect of *Zingiber officinale* and propolis on

- microorganisms and endotoxins in root canals. *J. Appl. Oral Sci.* 2013;21(1):25-31.
27. Rees LP, Minney SF, Plummer NT, Slater JH, Skyrme DA. A quantitative assessment of the antimicrobial activity of garlic (*Allium sativum*). *World J Microbiol Biotechnol* 1993;9:303-7.
 28. Reuter HD, Koch HP, Lawson DL. Therapeutic effects and applications of garlic and its preparations. In: *Garlic: The Science and Therapeutic applications of Allium sativum L. and Related species*, 2nd ed. (Koch H.P & Lawson D. L., eds.) 1996; 135-212.
 29. Suresh Pandey, Rhitu Shekhar, Rohit Paul, Manoj Hans, Amit Garg –comparative evaluation and effectiveness of A comparative evaluation and effectiveness of different antimicrobial herbal extracts as endodontic irrigants against enterococcus faecalis and candida albicans- an in-vitro study *University J Dent Scie* 2018; 4(2):75-81.
 30. Kandaswamy D, Venkateshbabu N, Gogulnath D, Kindo AJ. Dentinal tubule disinfection with 2% chlorhexidine gel, propolis, morinda citrifolia juice, 2% povidone iodine, and calcium hydroxide. *Int Endod J* 2010;43:419-423.
 31. Krithikadatta J, Indira R, Dorothykalyani AL. Disinfection of dentinal tubules with 2% chlorhexidine, 2% metronidazole, bioactive glass when compared with calcium hydroxide as intracanal medicaments. *J Endod* 2007;33:1473-1476.
 32. Gomes BP, Souza SF, Ferraz CC, Teixeira FB, Zaia AA, Valdrighi L, *et al.* Effectiveness of 2% chlorhexidine gel and calcium hydroxide against *Enterococcus faecalis* in bovine root dentin *in vitro*. *Int Endod J* 2003;36:267-275.
 33. Bhardwaj A, Ballal S, Velmurugan N. Comparative evaluation of the antimicrobial activity of natural extracts of Morinda citrifolia, papain and aloe vera (all in gel formulation), 2% chlorhexidine gel and calcium hydroxide, against *Enterococcus faecalis*: An *in vitro* study. *J Conserv Dent* 2012;15:293-297.